

***All lab tests going to GA Public Health Laboratory must be approved by CDPH/DPH Epidemiology**

GPHL Test Name	Description	Specimen Requirements	Test Method	Normal Value	Turn Around Time
Arbo Virus IgG	Detection of Arbovirus IgG antibody to determine immune status	<u>Specimen:</u> Serum <u>Container:</u> Serum Separator Tube (SST), or Red Top <u>Collection:</u> 4-6ml Red Top spin down, SST centrifuge and transfer serum to a transfer tube <u>Transport:</u> Overnight at room-temperature; Over 24 hours cold (2°-8°C)	IFA- CE EE SLE WE	Negative	7-14 Days
Arbo Virus IgM	Detection of Arbovirus IgM antibody to determine active and/or past infection	<u>Specimen:</u> Serum <u>Container:</u> Serum Separator Tube (SST), or Red Top <u>Collection:</u> 4-6ml Red Top spin down, SST centrifuge and transfer serum to a transfer tube <u>Transport:</u> Overnight at room-temperature; Over 24 hours cold (2°-8°C)	IFA- CE EE SLE WE	Negative	7-14 Days
Malaria Parasite	Speciation of malaria parasites by microscopic examination of Giemsa-stained blood smear	<u>Specimen:</u> Fiesma or Wright stained thick and thin blood smears plus EDTA whole blood <u>Container:</u> EDTA (lavender top) tube <u>Collection:</u> Draw successive blood samples in between shifts if possible <u>Transport:</u> Slides in slide hodler at room-temperature; EDTA blood in cold (2°-8°C)	Microscopy	No Parasites Found	1-3 Days
Malaria Parasites-PCR	DNA extraction and amplification by PCR	<u>Specimen:</u> Whole blood preferably less than 72 hrs <u>Container:</u> EDTA (Lavender top) tube <u>Collection:</u> At least 1 ml of blood by venipuncture <u>Transport:</u> Cold (2°- 8°C) (NOTE: Do not freeze)	Real-time PCR	No Parasites Found	1-2 Days
WNV IgG	Detection of West Nile Virus IgG antibody to determine immune status	<u>Specimen:</u> Serum <u>Container:</u> Serum Separator Tube (SST), or Red Top <u>Collection:</u> 4-6ml Red Top spin down, SST centrifuge and transfer serum to a transfer tube <u>Transport:</u> Overnight at room-temperature; Over 24 hours cold (2°-8°C)	EIA	Negative	5-7 Days
WNV IgG	Detection of West Nile Virus IgM antibody to determine active and/or past infection	<u>Specimen:</u> Serum <u>Container:</u> Serum Separator Tube (SST), or Red Top <u>Collection:</u> 4-6ml Red Top spin down, SST centrifuge and transfer serum to a transfer tube <u>Transport:</u> Overnight at room-temperature; Over 24 hours cold (2°-8°C)	EIA-serum EIA-CSF	Negative	5-7 Days
Zika Virus IgM	Detection of Zika Virus IgM antibody to determine active and/or past infection	<u>Specimen:</u> Serum <u>Container:</u> Red Top or Tiger top tube <u>Collection:</u> At least 1mL of blood Asymptomatic pregnant patients: IgM testing only and serum is drawn 2 to 12 weeks after return from travel.			

Zika Virus PCR	DNA extraction and amplification by PCR. Indicates current infection.	<p><u>Serum only:</u> <u>Specimen:</u> Serum <u>Container:</u> Red Top or Tiger top tube <u>Collection:</u> At least 1mL of blood PCR is being performed on serum samples collected ≤ 7 days after symptom onset and ELISA IgM is being performed on serum samples collected ≥ 4 days after symptoms onset (so sample collected between 4 and 7 days are receiving both PCR and IgM testing). Each serum test will need at least 1 ml of blood in a red top or tiger top tube.</p> <p><u>Serum and urine:</u> <u>Specimen:</u> Serum <u>Container:</u> Red Top or Tiger top tube <u>Collection:</u> At least 1mL of blood PCR is being performed on urine samples collected days 0-14 from symptom onset but the sample MUST have a paired serum sample also being tested for PCR. PCR must be run on serum samples that have a paired urine sample 0-14 days after illness onset. Each urine test will need at least 1 ml of urine in a sterile collection cup.</p> <p>Tape or parafilm the sample prior to shipping. Double bag the sample and place the GPLH form in the outer bag to prevent the form from being separated from the sample. If the sample is sent without a form, the lab will discard the sample.</p>			
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